GS01 0163 Analysis of Microarray Data

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Lecture 14: Biology: Functions and Networks

- GeneOntology
- GoMiner
- Regulatory networks and metabolic pathways

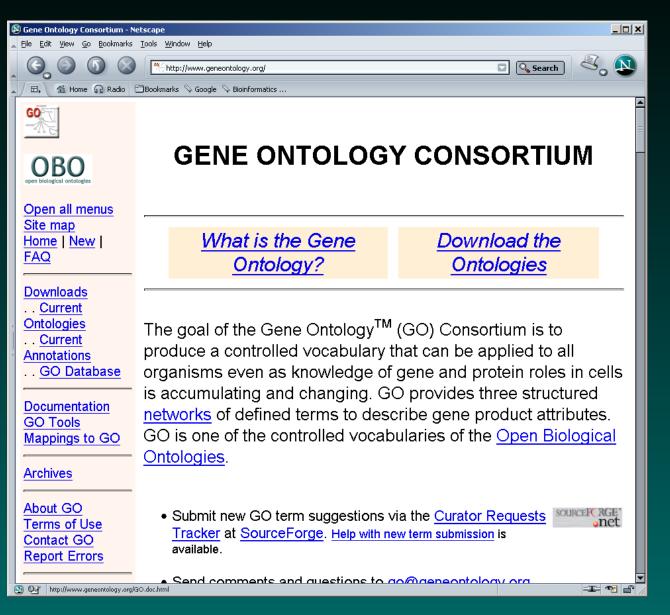
GeneOntology

GeneOntology (GO) uses controlled vocabularies to create a directed acyclic graph (DAG; a generalized tree) that describes the kinds of functions or properties that a gene might have.

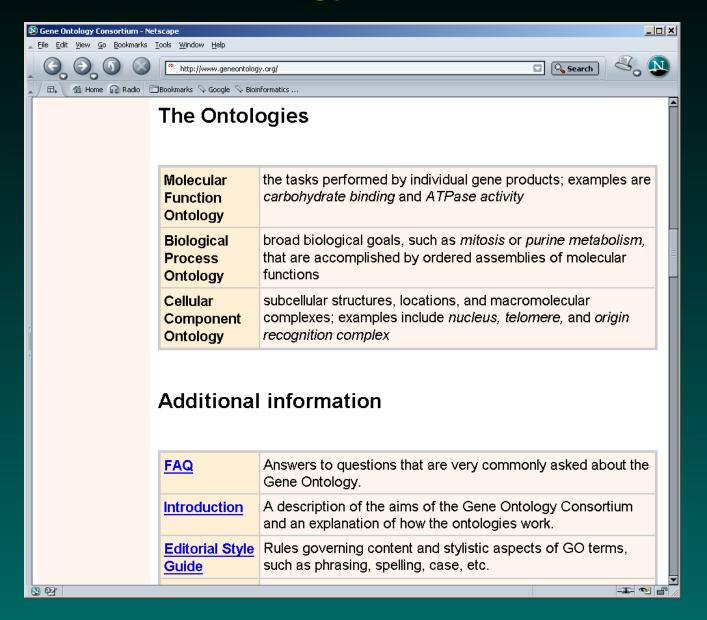
GeneOntology consists fundamentally of two pieces:

- 1. The DAG that describes functions and relations between them
- 2. Annotations that describe which genes actually have which functions

http://www.geneontology.org



GeneOntology: The top level

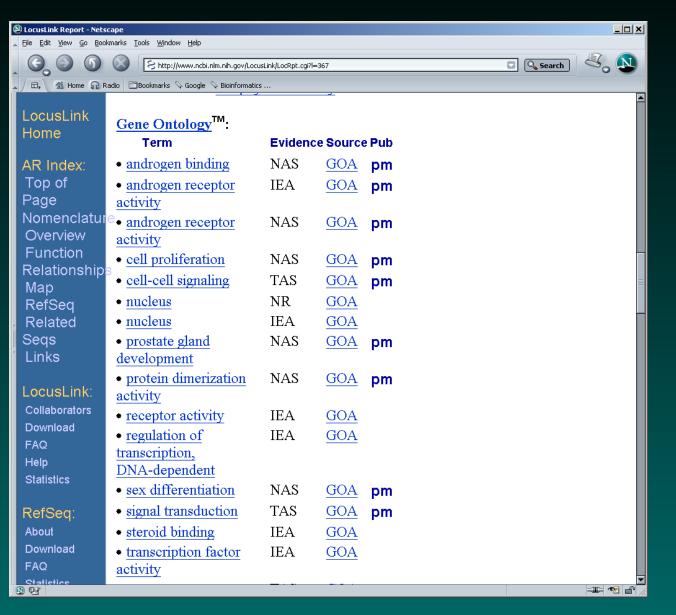


GeneOntology annotations in LocusLink

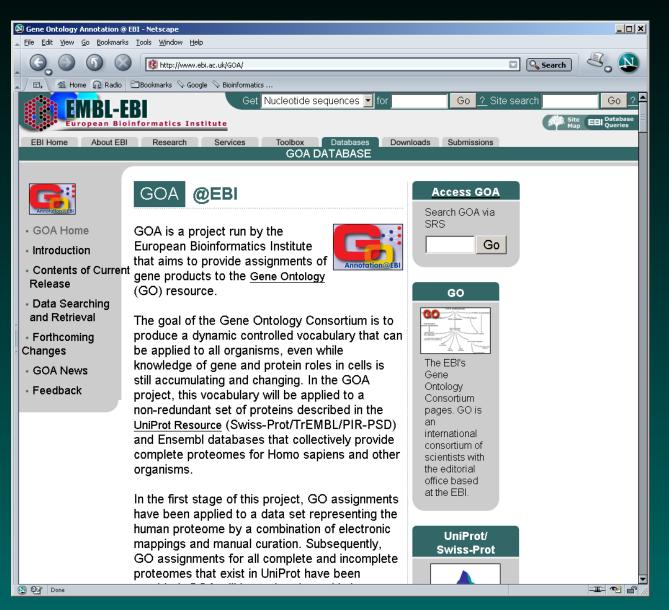
You can find the GeneOntology annotations for individual genes in LocusLink. For genes with known functions, the LocusLink page will conmtain a section titled "GeneOntology", which contains a list of the known functions for that gene.

Every GO annotation asserts that a specific gene has a specific function. As part of the design of GO, each assertion is itself annotated to explain the kinds of evidence the assertion is based on, as well as the organization or individual that supplied the annotation.

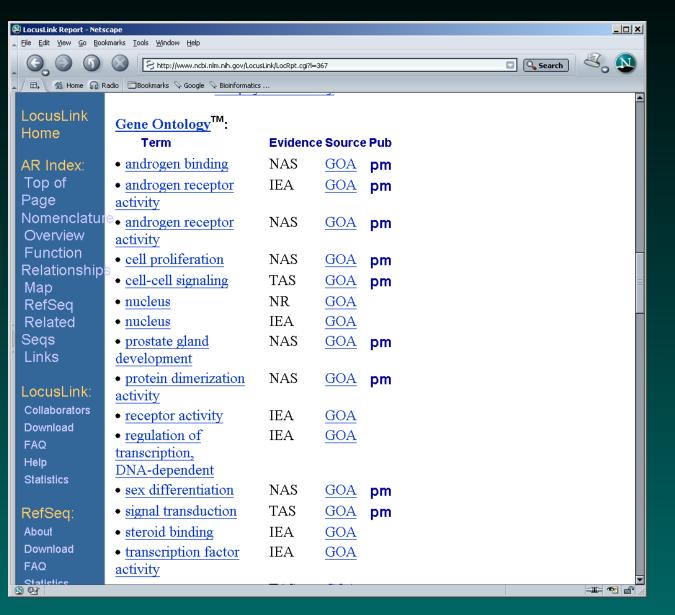
GO annotations of the androgen receptor



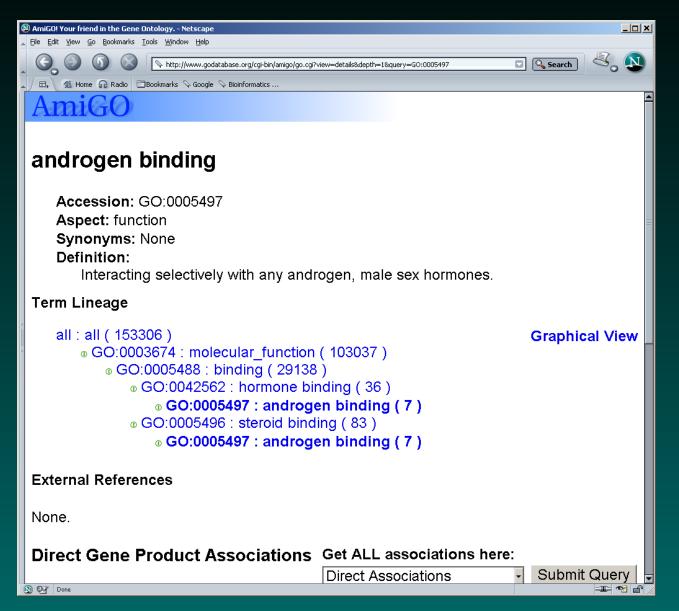
http://www.ebi.ac.uk/GOA/



GO annotations of the androgen receptor







GO browsing

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Edges are relationships

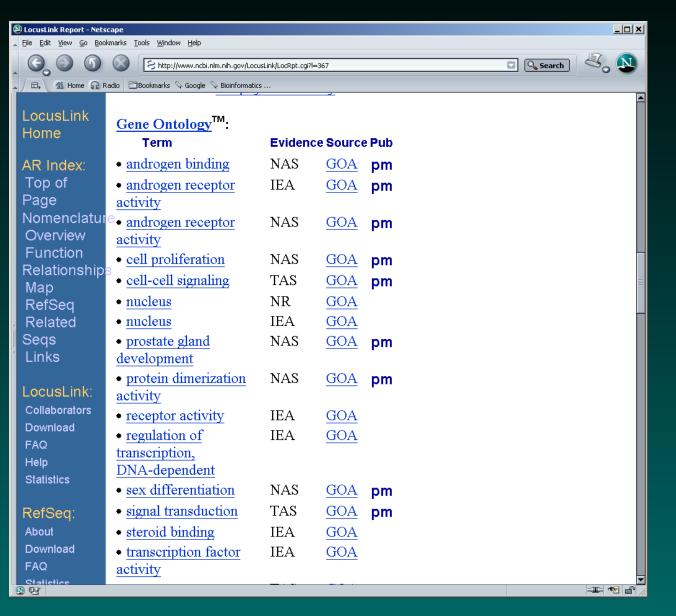
Edges in the DAG represent two kinds of relationships:

is_a : Used when the child node is a special case of the parent
 node. For example, hormone binding is_a kind of binding.

part_of : Used when the child node is a component of the parent
node. For example, a membrane is part_of a cell

Genes may be annotated into different levels of the hierarchy, depending on how detailed the evidence is. In general, a gene not only has the function corresponding to the node with direct annotation, but also has every property at parent nodes up through the hierarchy.

GO annotations of the androgen receptor



GeneOntology: Evidence Codes

- **IDA** : inferred from direct assay; indicates that the annotation is based on a paper describing an experiment that directly tested this function for this gene
- **TAS** : traceable author statement; based on a review article or textbook that includes references to the original experiments
- **IMP** : inferred from mutant phenotype; based on experiments involving mutations, knockouts, antisense, etc.
- **IPI** : inferred from physical interation; based on assays (like co-immunoprecipitation) that demonstrate physical interactions between the gene in question and other gene products

- **IGI** : inferred from genetic interaction; based on experiments (such as synthetic lethals, suppressors, functional complementation) that show a genetic interaction between the gene in question and another gene
- **ISS** : inferred from sequence or structure similarity; based on BLAST results that have been reviewed for accuracy by a curator
- **IEP** : inferred from expression pattern; based on Northerns, Westerns, or microarray experiments that reveal information about the timing or location of expression
- **NAS** : non-traceable author statement; statements in papers (abstract, introduction, discussion) that a curator cannot trace to another publication

IEA : inferred from electronic annotation; based on sequence similarity searches or database records that have not been reviewed by a curator

IC : inferred by curator; even though no direct evidence is available, the property can reasonably be inferred by the curator. For example, it is reasonable to infer from direct evidence of "transcription factor activity" that the gene product is found in the nucleus

ND : no biological data available; only used for annotations to "unknown"

NR : not recorded; used only for annotations created before curators started adding evidence codes

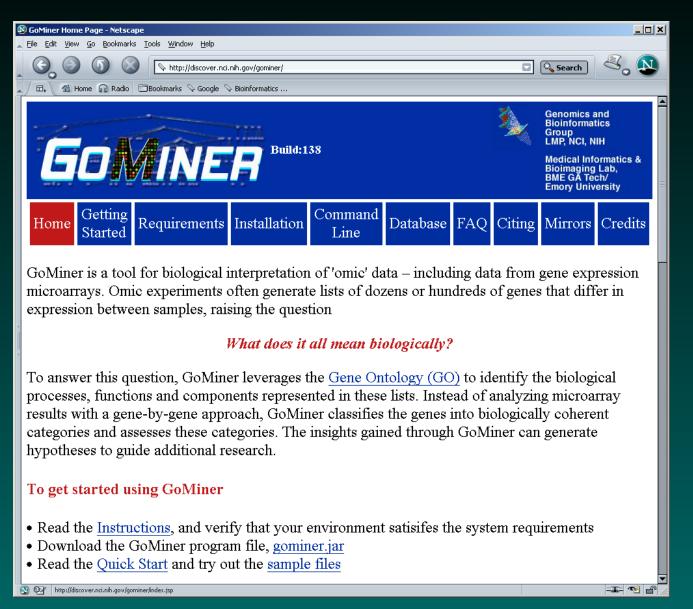
Quality of evidence

The evidence codes fall into a rough hierarchy indicating how strongly the annotation of function should be believed.

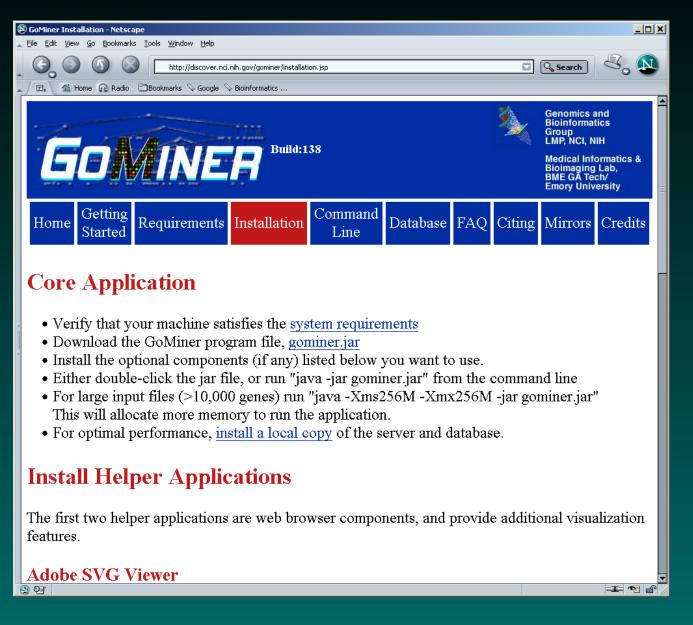
- 1. IDA, TAS
- 2. IMP, IPI, IGI
- 3. ISS, IEP
- 4. NAS
- 5. IEA

6. IC

GoMiner



http://discover.nci.nih.gov/gominer



GoMiner: Getting Started

You need a machine with

- Java 1.3 or higher
- Windows 98 or higher, Mac OS X or higher, Solaris, Linux, or FreeBSD
- High-speed ointernet access

Download the GoMiner Java code, install it, and double-click on it to start the program.

Then go to "File" - > "Load GO Terms" and click "OK".

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GoMiner: GO terms loaded

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GoMiner as GO browser

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Getting array data into GoMiner

- 1. Go to "Data Source" and select "UniProt (Hs)" to restrict to human gene annotations
- Need a file containing a list of all genes in the experiment, one HUGO symbol per line. Use the "Browse" button, and then click "Query Gene File" to load this information. This may take some time...
- Need a file containing a list of genes that changed. Can be one HUGO symbol per line. Optionally, you can include a second column with 1 (overexpressed) or -1 (underexpressed). Use "Browse" and "Query Changed Gene File" to load this data.

Note: GeneLink or Source can convert from various gene ids to HUGO symbols.

GoMiner with array gene list loaded

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File Edit Data Source Organisms View LookupSettings	
Input Genes Genes Mapped On GO	Summary View Selected Gene View
Input Gene Source Status top (1299)	Category Name P-Chng P-Undr P-Ovr Tot Chng Undr Ovr Category ID
YWHAE 143E UniProt 😨 🔺 🖻 🕕 biological_process (1245)	ATP-dependent hel., 1.0000 1.0000 1.0000 11 0 0 0 GO:00080 •
SFN 143S UniProt 😉 💹 🕀 🗊 behavior (8)	transcription elong 1.0000 1.0000 1.0000 1 0 0 0 GO:00080 20
PPP2R 2A5A UniProt 😉 🗉 🖬 🗊 biological_process unknown (27)	protein C-terminus 1.0000 1.0000 1.0000 2 0 0 0 GO:00080
PPP2R 2A5B UniProt 😉 🗉 🕀 🗊 cellular process (847)	microtubule binding 1.0000 1.0000 1.0000 3 0 0 0 GO:00080
PPP2R 2A5D UniProt 🞯 🗉 🖬 🕀 🛈 development (220)	regulation of heart r 1.0000 1.0000 1.0000 1 0 0 0 GO:00080
PPP2R 2A5E UniProt 😨 🛛 🕒 🗊 physiological process (1139)	circulation 1.0000 1.0000 1.0000 9 0 0 0 0 GO:00080
PPP2R 2A5G UniProt © GIGF2_HUMAN (IGF2) - (UniProt)	beta-catenin binding 1.0000 1.0000 1.0000 1 0 0 0 GO:00080
PPP2R 2AAA UniProt © GIGFA_HUMAN (IGF1) - (UniProt)	chemokine activity 1.0000 1.0000 1.0000 18 0 0 0 GO:00080
PPP2R 2AAB UniProt © - O43200 (TSHR) - (UniProt)	oligopeptide transp., 1.0000 1.0000 1.0000 1 0 0 0 GO:00151
PPP2R 2ABA UniProt © © PGH1_HUMAN (PTGS1) - (UniProt	peptide transporter 1.0000 1.0000 1.0000 2 0 0 0 GO:00151
PPP2R 2ABB UniProt © GPGH2_HUMAN (PTGS2) - (UniProt	Contemporaria and trans 1.0000 1.0000 1.0000 1 0 0 0 0 0 0 0
HLA-DMA 2DMA UniProt © GREL1_HUMAN (RLN1) - (UniProt)	acidic amino acid tr 1.0000 1.0000 1.0000 1 0 0 0 GO:00151
HLA-D 2DMB UniProt © GRLF_HUMAN (RLF) - (UniProt) 3	amino acid transpo 1.0000 1.0000 1.0000 2 0 0 0 0 GO:00151
HLA-DOA 2DOA UniProt 😉 🕢 🕀 🐨 cellular physiological process (50	
HLA-DRA 2DRA UniProt 😉 🛛 🕀 🐨 coagulation (16)	monosaccharide tr 1.0000 1.0000 1.0000 2 0 0 0 GO:00151
SH3BP2 3BP2 UniProt 😉 🛛 🔁 🐨 death (123)	carbohydrate trans 1.0000 1.0000 1.0000 3 0 0 0 GO:00151
SLC3A2 4F2_H UniProt G Decaging (2)	nitric oxide metabol 1.0000 1.0000 1.0000 4 0 0 0 GO:00462
A2M A2MG UniProt G D Cell death (122)	sodium ion transpo 1.0000 1.0000 1.0000 1 0 0 0 GO:00150
ACTN1 AAC1 UniProt G E cell aging (1)	hydrogen ion trans 1.0000 1.0000 1.0000 3 0 0 0 GO:00150
PRKAB1 AAKB UniProt 😉 🕀 🕀 😯 cytolysis (3)	monovalent inorga 1.0000 1.0000 1.0000 3 0 0 0 GO:00150
PRKAG1 AAKG UniProt 😉 👘 🖬 🗇 🙃 🗍 🕒	
ATBF1 ABF1 UniProt @ 🕀 🐨 apoptosis (120)	protein phosphatas 1.0000 1.0000 1.0000 3 0 0 0 GO:00150
ABL1 ABL1 UniProt 😨 🛛 👘 🗁 🗁 regulation of programme	
ABL2 ABL2 UniProt 🞯 🛛 🕀 🐨 extracellular structure organizati	ion and b glutathione disulfid 1.0000 1.0000 1.0000 1 0 0 0 GO:00150
ABR ABR_H UniProt @ 🛛 🕀 🐨 homeostasis (13)	peptide disulfide ox 1.0000 1.0000 1.0000 1 0 0 0 GO:00150
ACY1 ACY1 UniProt @ 🛛 🛨 🐨 metabolism (823)	disulfide oxidoredu 1.0000 1.0000 1.0000 4 0 0 0 GO:00150
ADAM17 AD17 UniProt 🞯 🛛 🛨 🐨 organismal physiological process	
ADA ADA_H UniProt 🞯 🕀 🐨 pathogenesis (3)	Cajal body 1.0000 1.0000 1 0 0 0 GO:00150
ADD3 ADDG UniProt @	(239) coreceptor activity 1.0000 1.0000 1.0000 6 0 0 0 GO:00150
ADH6 ADH6 UniProt 💿 🕀 🐨 response to stimulus (359)	glucuronosyltransf 1.0000 1.0000 2 0 0 0 GO:00150
ADK ADK_H UniProt 💿 🗄 🕀 🗊 secretion (2)	nuclear organizatio 1.0000 1.0000 25 0 0 0 GO:00069
AOX1 ADO_H UniProt 💿 🛛 🕀 🗊 regulation of biological process (403	
ADSS ADSS 💿 🗄 🐨 viral life cycle (8)	unfolded protein re 1.0000 1.0000 1.0000 1 0 0 0 GO:00069
SLC25A5 ADT2 UniProt 💿 🛛 🕀 🐨 cellular_component (1070)	alcohol catabolism 1.0000 1.0000 1.0000 4 0 0 0 GO:00461
MLLT2 AF4_H UniProt @ Indecular_function (1215)	response to unfold 1.0000 1.0000 1.0000 5 0 0 0 GC:00069
GLA AGAL UniProt 💿 🛛 🐨 obsolete_component	ER-nuclear signali 1.0000 1.0000 1.0000 1 0 0 0 GO:00069
ANGPT1 AGP1 UniProt 💿 🛛 🐨 obsolete_function	response to lipid hy 1.0000 1.0000 1.0000 1 0 0 0 GO:00069
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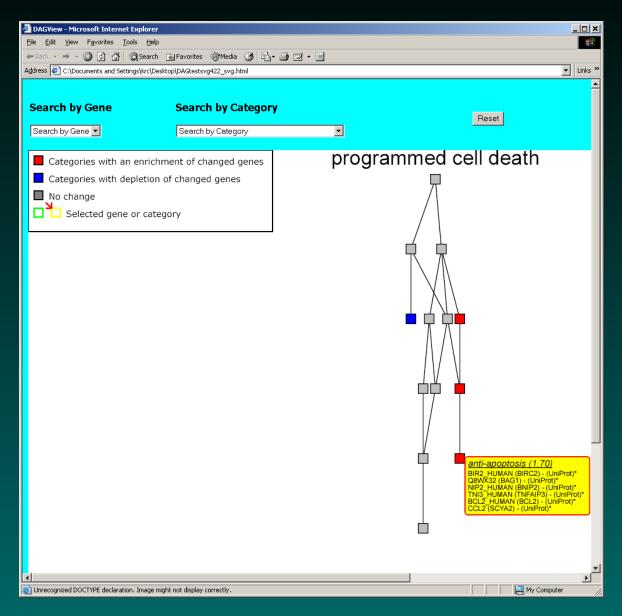
GoMiner with changed gene list loaded

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YWHAE 143E UniProt 🙆 🔺	1-1 biological_process (1245 1.03 p=0.17 1.01 p=0.48 1.02 p=0.17)	cytoplasmic seque 0.0002 0.0178 0.0260		2 2	GO:00429 A
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PPP2R 2A5A UniProt G	⊡ • • • • • • • • • • • • • • • • • • •	transcription factor 0.0002 0.0178 0.0260		2 2	GO:00429
PPP2R 2A5B UniProt G	⊕ @ development (220 0.96 p=0.62 1.12 p=0.35 1.04 p=0.43)	regulation of transc 0.0002 0.0178 0.0260		2 2	GO:00429
PPP2R 2A5D UniProt 🛛 😉	😑 🕕 physiological process (1139 1.08 p=0.04 1.05 p=0.11 1.06 p=0.01)	regulation of protei 0.0002 0.0178 0.0260		2 2	GO:00423
PPP2R 2A5E UniProt G	E - Cellular physiological process (568 1.02 p=0.48 0.99 p=0.57 1.01	regulation of nucleo 0.0002 0.0178 0.0260		2 2	GO:00468
PPP2R 2A5G UniProt G	⊕ ⊕ coagulation (16 1.10 p=0.61 0.90 p=0.69 0.99 p=0.62)	chemokine activity 0.0008 0.0782 0.0060		3 5	GO:00080
PPP2R 2AAA UniProt 💿	□ • • • • • • • • • • • • • • • • • • •	G-protein-coupled r 0.0008 0.0782 0.0060		3 5	GO:00016
PPP2R 2AAB UniProt 💿	□ • • • • • • • • • • • • • • • • • • •	chemokine recepto 0.0008 0.0782 0.0060		3 5	GO:00423
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PPP2R 2ABB UniProt @	🗆 🕕 🗊 programmed cell death (120 1.32 p=0.24 1.08 p=0.45 1.1	taxis 0.0012 0.0547 0.0112		5 7	GO:00423
HLA-DMA 2DMA UniProt 🛛 😉	⊕ ⊕ apoptosis (120 1.32 p=0.24 1.08 p=0.45 1.19 p=0.24)	response to wound 0.0015 0.0227 0.0296		9 10	GO:00096
HLA-D 2DMB UniProt 🛛 😉		response to chemi 0.0018 0.0814 0.0097		6 9	GO:00422
HLA-DOA 2DOA UniProt 🛛 🕲	⊕ • • • • • • • • • • • • • • • • •	response to pathog 0.0030 0.2972 0.0055		1 3	GO:00096
HLA-DRA 2DRA UniProt 🛛 🔸	⊞ - 🛈 metabolism (823 0.90 p=0.91 1.14 p=0.04 1.03 p=0.33)	regulation of transp 0.0030 0.0414 0.0593		2 2	GO:00510
SH3BP2 3BP2 UniProt 🛛 😉	🕀 🕕 organismal physiological process (254 1.87 p=0.00 0.91 p=0.71	immune response 0.0033 0.0002 0.4695		24 15	GO:00069
SLC3A2 4F2_H UniProt 💿	E regulation of physiological process (239 1.10 p=0.38 1.39 p=0.05 1	response to pest, p., 0.0036 0.0178 0.0743		13 13	GO:00096
A2M A2MG UniProt 💿	⊕ ⊕ response to stimulus (359 1.47 p=0.01 1.09 p=0.34 1.26 p=0.02)	extracellular space 0.0038 0.0039 0.2217		B 5	GO:00056
ACTN1 AAC1 UniProt 💿	⊕ - ⊕ regulation of biological process (403 1.18 p=0.18 1.25 p=0.06 1.22 p	protein threonine/tyr0.0063 0.0558 0.0794		2 2	GO:00047
PRKAB1 AAKB UniProt 💿	⊞- 🛈 viral life cycle (8 2.19 p=0.38 1.80 p=0.44 1.98 p=0.27)	MAP kinase kinase 0.0063 0.0558 0.0794		2 2	GO:00047
PRKAG1 AAKG UniProt 🛛 🕲	1-1 cellular_component (1070 0.97 p=0.78 0.97 p=0.78 0.97 p=0.84)	response to pathog 0.0063 0.3374 0.0092		1 3	GO:00428
ATBF1 ABF1 UniProt ©	1-1 molecular_function (1215 0.95 p=0.96 0.91 p=1.00 0.93 p=1.00)	antigen processing 0.0070 0.0001 1.0000		6 0	GO:00303
ABL1 ABL1 UniProt G	-O obsolete_component	antigen presentation 0.0070 0.0001 1.0000		6 0	GO:00198
ABL2 ABL2 UniProt ©	-O obsolete_function	MHC class II recept 0.0074 0.0024 0.5475		4 1	GO:00450
ABR ABR_H UniProt G	-O obsolete_process	response to extern 0.0075 0.0400 0.0743		12 13	GO:00096
ACY1 ACY1 UniProt ©		defense response 0.0088 0.0008 0.4993		24 16	GO:00069
ADAM17 AD17 UniProt ©		response to biotic s 0.0089 0.0013 0.4397		25 18	GO:00096
ADA ADA_H UniProt ©		inflammatory respo 0.0096 0.1695 0.0232		5 8	GO:00069
ADD3 ADDG UniProt G		innate immune res 0.0096 0.1695 0.0232		5 8	GO:00450
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MLLT2 AF4_H UniProt @		ion homeostasis 0.0114 1.0000 0.0008		0 5	GO:00508
GLA AGAL UniProt 😉		response to abiotic 0.0119 0.1597 0.0309		6 9	GO:00096
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GoMiner subgraphs

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SFN 143S UniProt 🙆 🧱	B • • • biological_process unknown (27 1.30 p=0.46 0.53 p=0.86 0.88 p=0.
PPP2R 2A5A UniProt 🙂	E • biological_process (847 0.99 p=0.58 0.97 p=0.69 0.98 p=0.67)
PPP2R 2A5B UniProt 🛛 🧿	B · ① development (220 0.96 p=0.62 1.12 p=0.35 1.04 p=0.43)
PPP2R 2A5D UniProt G	∃ · • • physiological process (1139 1.08 p=0.04 1.05 p=0.11 1.06 p=0.01)
PPP2R 2A5E UniProt G	⊕ • • • • • • • • • • • • • • • • •
PPP2R 2A5G UniProt ©	Coagulation (16 1.10 p=0.61 0.90 p=0.69 0.99 p=0.62) □ © programmed cell death (120 1.32 p=0.24 1.08 p=0.45 1.19 p=0.24) □ © non-second cell death (120 1.32 p=0.24 1.08 p=0.45 1.19 p=0.24)
PPP2R 2AAA UniProt G	□ • • death (123 1.28 p=0.26 1.17 p=0.34 1.22 p=0.20) □ • • • physiological process (1139 1.08 p=0.04 1.05 p=0.11 1.06 p=0.01) □ • • • cellular physiological process (568 1.02 p=0.48 0.99 p=0.57 1.00 p=0.51)
PPP2R 2AAB UniProt ©	Cell dealth (122 1.23 p=0.23 1.10 p=0.33 1.23 p=0.13)
PPP2R 2ABA UniProt ©	E C Cytolysis (3 0.00 p-1.00 4.0 r p-0.15 2.04 p-0.33)
PPP2R 2ABB UniProt ©	
HLA-DMA 2DMA UniProt @	$\square = 0$ coll death (122 1.20 p-0.25 1.10 p-0.33 1.23 p-0.40)
HLA-D 2DMB UniProt G	Ants_nowien (Antry_DAG of changed genes
HLA-DOA 2DOA UniProt ©	 ASP2_NOWAN (1P3) Evport DAC of changed genes to file
HLA-DRA 2DRA UniProt	A BAD HOWAN (BAD)
SH3BP2 3BP2 UniProt ©	- ↑ BCL2_HUMAN (BCL Export Genes By Category
SLC3A2 4F2_H UniProt ©	→ BIR2_HUMAN (BIRC2) - (UniProt)
A2M A2MG UniProt @	DAD1_HUMAN (DAD1) - (UniProt)
ACTN1 AAC1 UniProt @	DAP1_HUMAN (OAP) - (UniProt)
PRKAB1 AAKB UniProt ©	→ DPF2 (REG) - (UniProt)
PRKAG1 AAKG UniProt ©	→ ICE6_HUMAN (CASP6) - (UniProt)
ATBF1 ABF1 UniProt G	→ IKBA_HUMAN (NFKBIA) - (UniProt)
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GoMiner subgraphs



Intepreting GoMiner results

Enrichment is computed as

changed genes in category / total genes in category

changed genes on array / all genes on array

Statistical evidence of enrichment is based on a Fisher exact test.

Example

With the sample files supplied from GoMiner, they have an array with 1399 genes, of which 177 change expression. A total of 37 genes on the array are annotated to the biological process category of chemotaxis, and 12 of those 37 genes changed expression. So, we have a 2×2 contingency table that looks like

chemotaxis:	yes	no	all
Changed	12	165	177
Unchanged	25	1197	1222
Total	37	1362	1399

The p-value arising from a Fisher exact test with these values is 0.001226, which is the value reported by GoMiner for the enrichment of the chemotaxis category.

Intepreting GoMiner results

The p-values from the Fisher test are not corrected for multiple testing, but they should be since one is potentially looking at all GO categories. The categories are not independent, so it is not clear how to correct for multiple testing.

If one filters the gene list from the array before testing differential expression (for example, by removing low expressing or low variance genes), should those genes be included in the "query gene file" for the experiment?

The Fisher exact test is not completely appropriate, since genes can have multiple overlapping annoptations into the GO DAG.

No existing test exploits the quality of evidence for the GO annotations.

Regulatory networks and metabolic pathways

The harder question to be asked about the results of a microarray experiment is: what pathways or networks are changed?

Distinction:

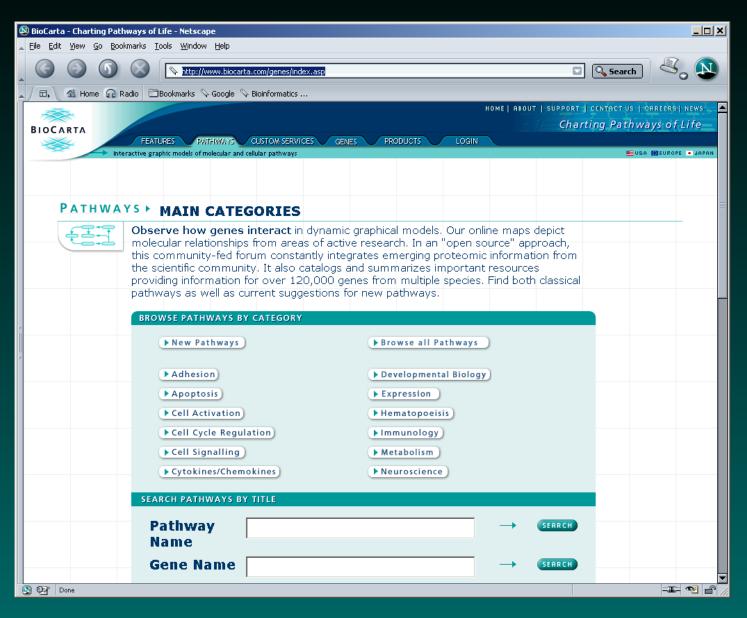
- Pathways typically refer to metabolic pathways (that process various small molecules) or signalling pathways that carry information from outside the cellinto the nucleus.
- Networks typically refer to interactons that control the expression level of various gene products.

Pathways are characterized by direct physical interactions betwen proteins. Networks are characterized by indirect interactions between genetic DNA or RNA or protein.

Pathway and netowrk resources

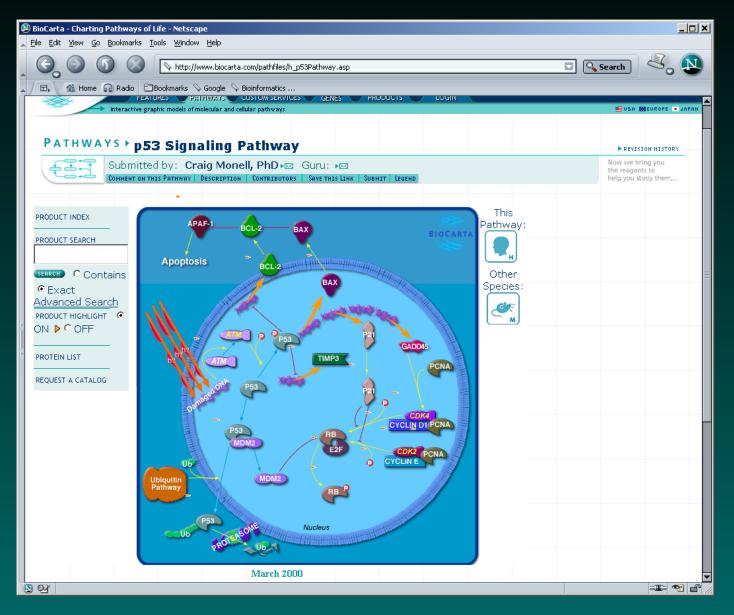
- KEGG (Kyoto Encyclopedia of Genes and Genomes) is a publically available tool, probably best used for metabolic pathways and secondarily for signalling pathways.
- PathArt (Jubilant Biosystems) is a commercial tool for investigating networks and pathways. Underlying network is based on curated interactions from the literature.
- Ingenuity Pathway Analysis (Ingenuity) is another curated commercial tool for investigating networks and pathways.

http://www.biocarta.com/genes/index.asp

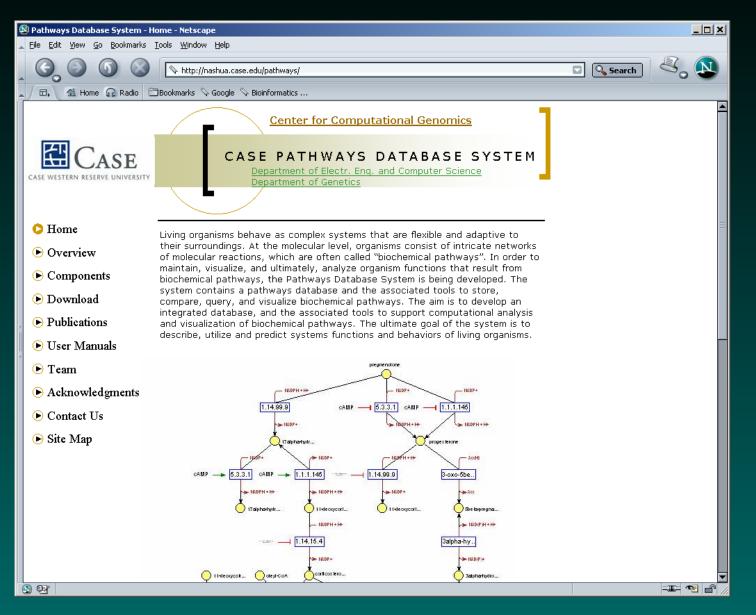


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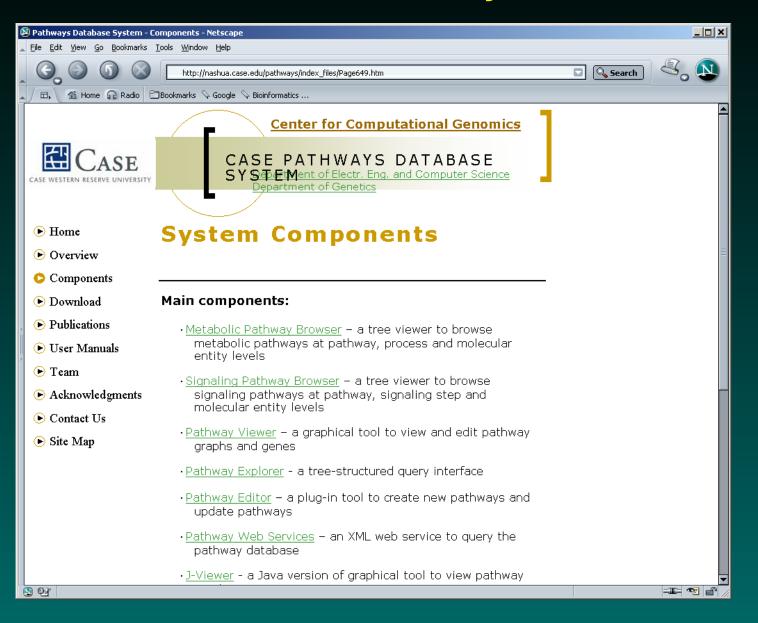
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http://nashua.case.edu/pathways



Case-Western Pathway Browser



Jubilant PathArt



PathArt[™] is a curated database of biomolecular interactions with tools for searching, analysis and visualization of data for use by microarray researchers and identification of potential drug targets. Pathway diagrams in PathArt[™] are dynamically generated from data in the database. PathArt[™] is accessible via any Java enabled browser and provides enterprise wide access to data stored in an Oracle database.

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Features

- Coverage of about 900 regulatory and signaling pathways across species.
- Browse pathways by organism, disease and other classifications.
- Coverage of protein-protein interaction.
- Information on knockout and mutagenesis studies.
- Search for pathways by specific genes.
- Coverage of 17 high priority diseases and disease responsive genes.
- Generate customized reports on genes and interactions of interest.
- Allows use of microarray expression data to search relevant pathways based on expression level. Allows use of Agilent and Affymetrix data.

Comprehensive information on all participating